Integumental Tyrosinase Activity in Amphibians

During the course of a study of the evolutionary aspects of skin tyrosinase activity¹, 2 amphibian species (Rana prpiens and Rana catesbiana) were found to possess the highest tyrosinase activity among the normal vertebrates studied and, in addition, were characterized by an unusual anatomic distribution of the enzyme. In order to characterize the nature of these features in other amphibian species, the present study has extended the original observations. The findings show that some urodeles possess the highest integumental tyrosinase activity levels among vertebrates.

Methods and materials. The Amphibia used (Table I) were decapitated, the dorsal and ventral skin areas immediately removed, trimmed and frozen (– 27 °C) for several weeks. At the time of assay, the frozen skin was sliced on dry ice and ground with chilled 0.1 M phosphate buffer (1:40) pH 6.8 as previously described 2. The assay procedures, substrates, methods of protein analysis, and statistical evaluation have been described 1,2 . Data are presented in the form, $\bar{x} \pm \sigma_{\bar{x}}$ wherever possible. The activities of the enzyme preparations are DOPA dependent, inhibited by 6 mM sodium diethyldithiocarbamate and are stable for at least 2 weeks at 0–4 °C. Hydrolysis (3 N HCl, 90 °C, 8 h) of the incubates did not significantly reduce the radioactivity, confirming the previously described findings 2 in other vertebrates.

Table I. Species utilized in the study of integumentary tyrosinase activity in Amphibia

Species	No. of animals used	Sex	Body weight (g), range	
(A) Urodela	-			
Amphiuma means (Congo eel)	2	3+9	3148, 6459	
Necturus maculosus (Mudpuppy)	3	13 + 29	94–149	
Ambystoma tigrinum (Tiger salamander)	3	b	31–41	
Diemictylus viridescens (Newt)	3 (1)*	ъ	2.0-2.3	
(B) Anura				
Rana pipiens (Leopard frog)	3	₫	45-53	
Rana catesbiana (Bull frog)	3	2♂ + 2♀	260-314	
Bujo marinus (Giant toad)	3	♂	163–173	

^a No. in parenthesis indicates the number of enzyme preparations used. Otherwise, the number of preparations is equal to the number of animals. ^b Data not available.

Table II. Comparison of in vitro tyrosinase activity in amphibian skin

Species	T.U./mg skin				Specific activ	Specific activity			
	Нь	p	S	P/S	Н	P	s	P/S	
Amphiuma i	means			-					
Dorsal Ventral	18,275 14,300	4,641 3,305	13,474 10,975	0.34 0.30	1,793 1,345	698 501	3,749 2,858	0.19 0.18	
Necturus ma	iculosus								
Dorsal Ventral	240 ± 21 126 ± 6	$\begin{array}{ccc} 167 \pm & 8 \\ 88 \pm & 3 \end{array}$	$\begin{array}{ccc} 65 \pm & 3 \\ 32 \pm & 1 \end{array}$	2.56 2.75	$\begin{array}{ccc} 22 \pm & 1 \\ 12 \pm & 0 \end{array}$	$\begin{array}{ccc} 23 \pm & 2 \\ 13 \pm & 1 \end{array}$	$\begin{array}{ccc} 19 \pm & 2 \\ 9 \pm & 0 \end{array}$	1.21 1.44	
Ambystoma i	tigrinum								
Dorsal Ventral	$2,115 \pm 76$ $16,646 \pm 231$	$1,680 \pm 10$ $9,639 \pm 10$	439 ± 8 $6,956 \pm 31$	3.82 1.38	84 ± 4 626 ± 14	$\begin{array}{ccc} 105 \pm & 5 \\ 526 \pm & 6 \end{array}$	$\begin{array}{c} 46 \pm 3 \\ 842 \pm 9 \end{array}$	2.28 0.62	
Diemictylus	viridescens								
Dorsal Ventral	16,238 9,085	15,062 5,892	2,720 3,213	5.53 1.83	780 464	1,584 585	249 345	6.36 1.70	
Rana pipien	ıs								
Dorsal Ventral	$1,112 \pm 60$ $1,510 \pm 13$	866 ± 27 $1,097 \pm 67$	250 ± 3 492 ± 5	3.46 2.22	$\begin{array}{ccc} 47 \pm & 2 \\ 52 \pm & 2 \end{array}$	$\begin{array}{ccc} 61 \pm & 2 \\ 47 \pm & 1 \end{array}$	$\begin{array}{ccc} 25 \pm & 1 \\ 66 \pm & 2 \end{array}$	2.44 0.71	
Rana catesbi	iana								
Dorsal Ventral	847 ± 49 $1,114 \pm 20$	$\begin{array}{c} 396 \pm 8 \\ 513 \pm 6 \end{array}$	471 ± 5 592 ± 5	0.84 0.86	$\begin{array}{ccc} 108 \pm & 8 \\ 137 \pm & 3 \end{array}$	84 ± 3 94 ± 2	$\begin{array}{c} 155 \pm & 3 \\ 219 \pm & 4 \end{array}$	0.54 0.43	
Bujo marini	us								
Dorsal Ventral	$1,400 \pm 36$ $1,632 \pm 17$	$357 \pm 22 \\ 595 \pm 18$	$1,061 \pm 12$ $1,054 \pm 26$	0.33 0.56	226 ± 13 229 ± 6	$\begin{array}{c} 112\pm10 \\ 146\pm6 \end{array}$	$348 \pm 18 \\ 351 \pm 5$	0.32 0.42	

^{*1} T.U. (tyrosinase unit) is defined as the amount of tyrosinase activity required to convert 1 picomole of L-tyrosine to melanin under the conditions of the described assay during a 16 h incubation period at 30 °C. Specific activity is defined as the number of T.U./ μ g protein nitrogen in the enzyme preparation. b H, homogenate; P, particulate fraction; S, soluble fraction.

¹ Y. M. CHEN and W. CHAVIN, Adv. Biol. Skin 8, 253 (1967).

Y. M. CHEN and W. CHAVIN, Analyt. Biochem. 13, 234 (1965).

Table III. In vitro tyrosine carboxyl group incorporation into melanin by amphibian skin tyrosinase

Species	Skin region	Tyrosine carboxyl group incorporation (%)a				
		Нь	P	S		
Amphiuma means	Dorsal Ventral	22.6 19.3	25.5 24.1	21.9 17.0		
Necturus maculosus	Dorsal Ventral	14.9 ± 0.5 13.5 ± 0.3	15.3 ± 0.4 13.5 ± 0.1	15.8 ± 0.3 15.8 ± 0.3		
Ambystoma tigrinum	Dorsal Ventral	$10.0 \pm 0.3 \\ 3.6 \pm 0.1$	$9.7 \pm 0.3 \\ 3.6 \pm 0.1$	$8.5 \pm 0.5 \\ 3.7 \pm 0.2$		
Diemictylus viridescens	Dorsal Ventral	11.4 15.8	11.3 15.8	10.3 15.8		
Rana pipiens	Dorsal Ventral	$37.3 \pm 1.0 \\ 38.2 \pm 1.1$	38.4 ± 0.6 39.0 ± 0.5	$37.4 \pm 1.9 \\ 36.9 \pm 0.8$		
Rana catesbiana	Dorsal Ventral	$21.9 \pm 1.0 \\ 23.5 \pm 0.6$	$18.5 \pm 0.6 \ 24.2 \pm 0.4$	21.3 ± 0.9 21.8 ± 0.5		
Bufo marinus	Dorsal Ventral	38.9 ± 1.7 34.0 ± 1.1	$31.6 \pm 1.1 \\ 31.8 \pm 1.1$	$41.0 \pm 0.9 \\ 35.3 \pm 1.0$		

^{*} Expressed in % of total L-tyrosine converted. b See footnote b, Table II.

Results and discussion. Six amphibian species showed very high integumental tyrosinase activities when compared with previously studied vertebrates1. As sex differences in tyrosinase activity were not discernible, all data for a species were consolidated. The enzymatic activity in both the dorsal and ventral skin of Amphiuma and Diemictylus and that in the ventral skin of Ambystoma (Table II) was greater than that occurring in any other normal vertebrate integument studied to date and approached or exceeded the enzymatic activity levels in the vertebrate melanomas previously studied. The very high specific activities in these amphibians indicate the relatively high concentration of tyrosinase in the protein pool of the skin. In addition, it is also possible that a highly effective tyrosinase activator(s) or that a less effective tyrosinase inhibitor(s) is present.

The integumental tyrosinase activity levels among the amphibians were, in descending order, Amphiuma, Diemictylus, Ambystoma, Bufo, R. pipiens, R. catesbiana and Necturus. In the dorsal skin, the highest activity present in Amphiuma was 76-fold higher than that present in the lowest, Necturus. In the ventral skin, the highest activity present, in Ambystoma, was 132-fold higher than that present in the lowest, Necturus. While the tyrosinase activity in the dorsal skin was 1.90-, 1.79and 1.28-fold greater than that in the corresponding ventral skin of Necturus, Diemictylus and Amphiuma, respectively. The tyrosinase activity in the ventral skin was 7.87-, 1.36-, 1.32- and 1.17-fold greater than that in the corresponding dorsal skin in Ambystoma, R. pipiens, R. catesbiana and Bufo, respectively. In R. pipiens and R. catesbiana the unusual anatomic distribution of tyrosinase activity and the possible mechanism involved have been discussed previously3. The findings in regard to Buto and Ambystoma may be explained similarly.

The subcellular distribution study of tyrosinase revealed that the enzyme was present in both the soluble and particulate fractions in all species (Table II). However, the enzymatic activity was greater in the particulate fraction of *Diemictylus*, *Ambystoma*, *R. pipiens* and *Necturus* skin. The highest ratio of particulate activity to soluble activity occurred in the dorsal skin of *Diemictylus* and the lowest in the ventral skin of *Amphiuma*.

The incorporation of tyrosine without decarboxylation in % of total L-tyrosine converted, ranged from 3.6-41.0%

(Table III). The highest occurred in Bufo and R. pipiens and the lowest in Ambystoma. However, Diemictylus and Necturus also showed a comparatively low carboxyl incorporation. The incorporation of tyrosine carboxyl groups into melanin supports the attraction hypothesis in melanogenesis as proposed previously 2,4 .

Summary. The anatomic and subcellular distribution of integumental tyrosinase activity and the tyrosine carboxyl group incorporation into melanin in 7 species of amphibia were investigated. Necturus maculosus, Diemictylus viridiscens and Amphiuma means showed higher tyrosinase activity in the dorsal skin compared to the ventral skin. However, Ambystoma tigrinum, R. pipiens, R. catesbiana and Bufo marinus showed the unusual feature of a higher enzymatic activity in the light ventral skin when compared to the dark dorsal. The integumental tyrosinase activity was present in both subcellular fractions, soluble and particulate, in all species. The particulate fraction of Diemictylus, Ambystoma, R. pipiens and Necturus was enzymatically more active. In the other species studied, the soluble fraction contained the greater enzymatic activity. The highest normal vertebrate integumental tyrosinase activity and specific activity reported to date, occur in the Amphibia. Tyrosine carboxyl incorporation into melanin ranged from 3.6-41.0%. Amphibia appear to be unusually interesting experimental animals in pigmentation studies 5.

Résumé. L'analyse radiométrique de tyrosinase intégumentale a démontré que les amphibiens sont les vertébrés les plus intéressants dans les études de mélanogenèse.

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³ Y. M. CHEN and W. CHAVIN, in manuscript (1967).

⁴ Y. M. CHEN and W. CHAVIN, Nature 210, 35 (1966).

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